

Ghalichi et al, **Health Promotion Perspectives**, 2022, 12(2), 122-130 doi: 10.34172/hpp.2022.16 https://hpp.tbzmed.ac.ir

Systematic Review





Vanadium and biomarkers of inflammation and oxidative stress in diabetes: A systematic review of animal studies

Faezeh Ghalichi¹⁰, Alireza Ostadrahimi², Maryam Saghafi-Asl^{3*0}

¹Student Research Committee, Tabriz University of Medical Sciences, Tabriz, Iran

²Department of Clinical Nutrition, Faculty of Nutrition and Food Sciences, Tabriz University of Medical Sciences, Tabriz, Iran ³Nutrition Research Center, Drug Applied Research Center, Department of Clinical Nutrition, Faculty of Nutrition & Food Sciences, Tabriz University of Medical Sciences, Tabriz, Iran

ARTICLE INFO

Article History: Received: 4 Feb. 2022 Accepted: 24 July 2022 ePublished: 20 Aug. 2022

Keywords:

Systematic review, Animals, Diabetes mellitus, Inflammation, Oxidative stress

*Corresponding Author: Maryam Saghafi-Asl, Email: saghafiaslm@gmail. com

Abstract

Background: Oxidative stress has a significant role in the commencement and development of hyperglycemia. Vanadium, as a transitional metal with redox properties, enters the redox process, produces free radicals, and distracts the pro-antioxidant balance. The present animal systematic review aimed to assess the effect of vanadium supplementation on inflammation and oxidative stress biomarkers in diabetes-induced animals.

Methods: A systematic search was conducted using the PubMed, Scopus, and web of science databases from 1990 to 2021, according to the inclusion and exclusion criteria. The search strategy was based on the guidelines for systematic review of animal experiments and Preferred Reporting Items for Systematic Reviews (PRISMA). Criteria for eligibility were animal-based studies, evaluating the therapeutic effects of vanadium on inflammatory and oxidative stress biomarkers in diabetes. The Systematic Review Centre for Laboratory Animal Experimentation (SYRCLE) tool was used for assessing the methodological quality of included studies.

Results: In the present study, 341 articles were evaluated out of which 42 studies were eligible for inclusion. The majority of the studies confirmed the advantageous properties of vanadium on inflammatory and oxidative stress biomarkers. A minor risk of bias was reported, based on the SYRCLE's tool.

Conclusion: According to the findings, well-designed clinical trials are warranted to assess the long-lasting effects of various vanadium compounds on inflammatory and oxidative stress biomarkers.

Introduction

Type 2 diabetes mellitus (T2DM), represents nearly 95% of all cases of DM and is characterized by insulin resistance or a decline in β -cells' ability to secrete insulin.¹ In chronic hyperglycemia, glucose auto-oxidation leads to abundant production of oxygen-free radicals in the mitochondria due to major oxygen usage, high redox reactions, mitochondrial-fission state, and failure of the antioxidant defense system.2-4 Nevertheless, oxidative stress has a significant role in the commencement and ongoing of hyperglycemia, as well. In general, the inequity of reactive oxygen species (ROS) production and elimination is described as oxidative stress.1 ROS production leads to the impairment of nuclear deoxyribonucleic acid (DNA). Additionally, it stimulates nuclear poly (ADP-ribose) polymerase (PARP), prevents glyceraldehyde-3-phosphate dehydrogenase (GAPDH) activity, and shunts primary glycolytic substrates into pathogenic signaling pathways

via the activation of I) the polyol, II) Protein kinase C (PKC), and III) glycation end-products (AGE) pathways.⁵

The signaling pathways mentioned above augment ROS formation and stimulate inflammation. The polyol pathway intensifies ROS production using nicotinamide adenine dinucleotide phosphate (NADPH) and glutathione (GSH), and aggregating consequent nicotinamide adenine dinucleotide (NADH) oxidation. In addition, hyperglycemia reinforces inflammatory cytokines, including tumor necrosis factor-alpha (TNF-α) and nuclear factor kappa-light-chain-enhancer of activated B-cells (NF-κB) expression. Inhibition of GAPDH leads to dihydroxyacetone phosphate construction, as well as PKC and AGE increase. Such events consequently induce NADPH oxidase and the expression of inflammatory factors and decline endothelial nitric oxide synthase stimulation. Also, PKC stimulates insulin resistance via preventing downstream expression of phosphatidylinositol

^{© 2022} The Author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (http:// creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

3-kinase (PI3K)/protein kinase B (Akt) signaling pathway (PI3K-Akt).^{4,5}

Cellular ROS concentration is detected by the production and clearance rate of ROS. Antioxidant enzymes such as superoxide dismutase (SOD), catalase (CAT), glutathione reductase (GR), and glutathione peroxidase (GPx) are substrates that scavenge free radicals or inhibit their conversion to toxic derivatives.⁶ Thus, modulating these enzymes protect the cellular antioxidant system from oxidative stress.

Since 1970s, the insulin-mimetic or insulin-enhancing properties of vanadium have been discussed and it has been considered as a therapeutic agent against diabetes.⁷ Vanadium, as a transitional metal with redox properties, enters the redox process, produces free radicals, and distracts the pro-antioxidant balance in the cell. Vanadium is a scavenger of superoxide radicals, and declines antioxidant enzymes such as SOD, GPx, CAT, and GR in erythrocytes,⁸⁻¹¹ liver,¹²⁻²² kidneys,^{14,19,20,22} heart,²² brain,^{22,23} pancreas^{16,20,24} and testes²⁵ of rats. Based on the results of multiple studies, vanadium complexes increase the action of GPx and demolish the effect of ROS in diabetic-induced rats.^{6,8,22}

Vanadium prevents protein tyrosine phosphatase activity and helps glucose transporter 4 translocation.²⁶ Redox regulation inhibits PTP-1B activation. Due to the insulin-stimulating properties of NAD(P)H oxidase homolog Nox 4, it modulates H2O2 and plays an essential role in insulin signaling via modulating PTP-1B transcription.²⁷ The complications of diabetes are directly associated with oxidative stress; hence, substrates reducing oxidative stress, are also beneficial for the complications of diabetes.²⁸

The beneficial effects of vanadium in declining hyperglycemia have already been reported in experimental and clinical trials.^{29,30} However, the objective of the current animal-based systematic review was to put together experimental evidence to present a thorough assessment of the effects of vanadium administration on inflammatory and oxidative stress biomarkers in diabetes-induced animals.

Material and Methods

Search strategy

The Preferred Reporting Items for Systematic Reviews (PRISMA) was implicated in this systematic review.³¹

PubMed, Scopus, and Web of Science databases were used for searching animal-based studies evaluating the effect of vanadium administration on inflammatory and oxidative stress biomarkers among diabetic animals from 1990 to 2021 (Table 1).³²⁻³⁴ Language restriction was not considered during the search strategy.

Inclusion criteria

Eligible studies for including in this systematic review obeyed the **PICOS** criteria, as below³⁵:

- Participants: Diabetes-induced laboratory animals.
- Interventions: Vanadium administration.
- Comparisons: Diabetic control animals, consuming a regular diet.
- Outcomes: Measuring inflammatory and oxidative stress biomarkers.
- Study design: Animal studies assessing the effect of vanadium administration in diabetic-induced animals.

Exclusion criteria

Studies assessing the effects of vanadium compounds on glycemic markers and lipid profile were excluded in this systematic review. Also, studies with invasive surgical procedures or certain diets were excluded.

Study selection

Animal studies were screened individually by two investigators, according to the inclusion and exclusion criteria. At first, the titles and abstracts of selected studies were assessed; afterwards, the full texts were read carefully. In the end, the papers were monitored for final detection. Disagreements regarding selecting certain studies for inclusion were determined by discussion among investigators.

Data extraction

A pre-standardized data extraction form was independently administered by two authors for extracting data. In the end, a third author was responsible for rechecking extracted data.

Assessment of methodological quality

The Systematic Review Centre for Laboratory Animal Experimentation (SYRCLE)'s Risk of Bias (RoB) tool³⁶ was used for evaluating the methodological quality and risk of

Table 1. Search strategy according to database filters

Database	Search items
PubMed	(("Vanadium"[Mesh] OR "Vanadium Compounds"[Mesh]) OR (vanadium[Title/Abstract])) AND (((((("Diabetes Mellitus, Type 2"[Mesh]) OR "Obesity"[Mesh]) OR "Glucose Intolerance"[Mesh]) OR ("Diabetes Mellitus"[Mesh] OR "Diabetes, Gestational"[Mesh] OR "Diabetes Mellitus, Type 1"[Mesh] OR "Diabetes Mellitus, Experimental"[Mesh]) OR "Insulin"[Mesh]) OR "Glycated Hemoglobin A"[Mesh]) OR (((((((Giabetes[Title/Abstract])) OR (obesity[Title/Abstract])) OR ("glucose intolerance"[Title/Abstract])) OR (Insulin[Title/Abstract])) OR ("Glycated hemoglobin A"[Title/Abstract])) OR (HbA1c[Title/Abstract])) OR (prediabetes[Title/Abstract])) OR (overweight[Title/Abstract])))
Scopus	((TITLE-ABS-KEY(Vanadium))) AND ((TITLE-ABS-KEY(Diabetes) OR TITLE-ABS-KEY (Obesity) OR TITLE-ABS-KEY (Overweight) OR TITLE-ABS-KEY (#Glucose Intolerance") OR TITLE-ABS-KEY (Insulin) OR TITLE-ABS-KEY (#Glucose Intolerance") OR TITLE-ABS-KEY (#Glucose Intolera
Web of Science	((Vanadium)) AND ((Diabetes) OR (Obesity) OR (Overweight) OR ("Glucose Intolerance") OR (Insulin) OR ("Glycated hemoglobin A") OR (HBA1C) OR Prediabetes))

bias of studies included, according to the Cochrane RoB tool. The SYRCLE's RoB tool assesses 10 items, including selection bias, performance bias, detection bias, attrition bias, reporting bias, and other biases.

Outcomes

Outcomes extracted from the included studies for evaluating the beneficial effects of vanadium were: (1) Inflammatory biomarkers such as TNF- α , interleukin 6 (II-6), high-sensitivity C-reactive protein (hs-CRP); (2) oxidative stress biomarkers including GSH, SOD, CAT, GPx, glutathione S-transferase (GST), and GR.

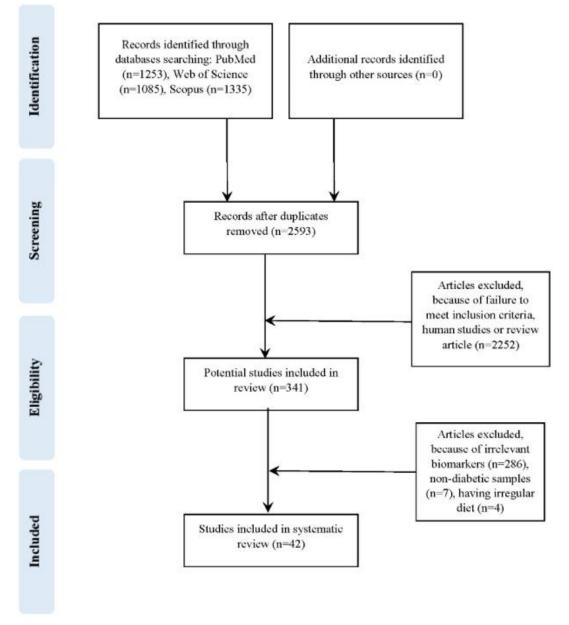
Results

Identification of relevant studies

During the electronic search, 2593 potentially eligible studies were identified. Then, reviewing the title and abstract of studies resulted in the exclusion of 2252 studies, due to not fulfilling the inclusion criteria, or being randomized controlled trials or review articles. Afterward, 341 full-text papers were further reviewed. Eventually, 42 studies were eligible for inclusion. A flow diagram outlining the selection of final papers can be observed in Figure 1. Among the 42 studies included, 40 studies reported the beneficial therapeutic effects of vanadium on the enzymatic activity of inflammatory and oxidative stress biomarkers in diabetes-induced animal studies.

Characteristics of studies included in the analysis

Table S1 summarizes the characteristics of animal studies included. The studies' publication dates ranged from August 1990³⁷ to November 2021.³⁸ In 24 out of the included studies,^{12,16-18,20,21,39-56} randomization was reported. Sample size was from 20⁵⁷ to 90² animals. Streptozotocin^{2,12-14,16-21,37-42,44,45,48-61} or alloxan monohydrate^{6,8,11,22,24,46,47,62,63} were used for inducing





diabetes, except for one study43 that used high-fat lowsucrose diet for inducing insulin resistance. All of the studies were accomplished on rodents. In 8 studies, 38,40,41,44,58,61 Sprague-Dawley rats were used, while in the rest, Swiss Albino of Wistar strain rats were investigated. Additionally, 3 studies^{2,16,43} were on mice species. Rat's mean weight was 175 g and mice's mean weight was 26.5 g. Sources of vanadium consumed for administration were vanadyl sulfate (20 studies),^{2,12,17,18,21,39-42,44,48-51,53,54,57,58,60} sodium orthovanadate (10 studies),^{6,8,11,19,22,37,46,62-64} oxovanadium (IV) complex (1 study),² vanadium pentoxide (2 studies),²⁰ Na[(O2)2(2,2'-bpy)] • 8 H2O vanadium complexes (1 study),¹³ vanadyl(IV)-ascorbate (VOAsc) complex (1 study),⁴³ dioxidovanadium cis-[VO2(obz)py]) complex (1 study),38 NaVO3 (3 studies),21,47 vanadium-3-hydroxy flavone complex (1 study),⁵⁹ macrocyclic binuclear oxovanadium (IV) complex (MBOV) (1 study),14 bis(maltolato)oxovanadium (IV) (2 studies),16,45 dioxidovanadium (1 study),61 vanadium citrate (1 study),24 V3dipic-Cl²¹ and oxovanadium(IV) chelate [VOL] (1 study).⁵¹ Method of administration was either by dissolving drinking water, ^{6,8,11,16,17,19,21,22,24,37,40,41,444,47,57-59,62,63,65} into gavage,^{12-14,18,20,38,39,42,43,48-54,61} or intraperitoneal injection.⁶⁰ Duration of the interventions ranged from 15 days up to 60 days. Measured outcomes were the enzymatic activity of inflammatory biomarkers such as TNF-a, Il-6, hs-CRP, caspase 3, as well as oxidative stress biomarkers including GSH, SOD, GPx, GST, and GR.

Quality assessment

Randomization was reported in 24 of the included studies.^{12,16-18,20,21,39-56} Animals were similar in age and weight and were kept in controlled and similar conditions. Figure 2 illustrates the methodological quality assessment of studies and Figure 3 shows the risk of bias of each item, as percentages.

Overview of outcome measures

Among the included studies, 18 studies assessed the effectiveness of vanadium on GSH. In all studies, GSH level was significantly increased, except for one study,⁴⁶ which observed no significant differences. Vanadyl sulfate supplementation significantly enhanced GSH levels in diabetes-induced animals.^{12,17,18,39,42,49,50,52} Other vanadium complexes such as oxovanadium (IV), Na[(O2)2(2,2'-bpy)] • 8 H2O, Na[VO(O2)2(1,10'-phen)] • 5 H2O, [VO(SO4)(1,10'-phen)] • 2 H2O, vanadyl (IV)-ascorbate (VOAsc), vanadium-3-hydroxy flavone, macrocyclic binuclear oxovanadium (IV) complex (MBOV), sodium orthovanadate, vanadium citrate, vanadium pentoxide, oxovanadium (IV) chelate also augmented GSH level.^{2,13,14,24,37,43,51,59}

Among the studies included, 27 studies analyzed the effectiveness of vanadium on GR, GST, and GPx levels. In one study, vanadium administration was inefficient in enhancing GPx level,⁴⁶ and in five studies, vanadium administration significantly declined GPx levels.^{16,42,53,54,63}

However, in the remaining of the studies, GPx level was significantly enhanced.^{6,8,11,14,19-22,24,37,38,43,48,51,59,61} Vanadium administration significantly increased GST level in 7 studies.^{8,11,17,48,50,57,62} In one study,¹¹ no significant changes were observed. Vanadium administration significantly enhanced GR level in 4 studies.^{8,11,19,24} No significant changes were observed in one study,⁴² but a significant decline was observed in two studies.^{53,54}

In addition, 27 studies assessed the effect of vanadium on SOD level out of which 20 observed significant enhancement. Also, in 3 studies, significant alterations were not observed.^{13,45,46} In five studies, vanadium treatment declined SOD level.^{16,42,47,53,54} Vanadyl sulfate supplementation could also significantly augment SOD level.^{2,21,41,42,44,48,50,53,54,60} Other compounds such as oxovanadium (IV) complex, sodium orthovanadate, [VO(SO4)(1,10'-phen)] • 2 H2O, [VO(SO4) (2,2'-bpy)] • H2O, vanadyl(IV)-ascorbate, Dioxidovanadium (V) complex, vanadium-3-hydroxy flavone complex, macrocyclic binuclear oxovanadium (IV) complex, bis(maltolato)oxovanadium IV (BMOV), vanadium citrate, vanadium pentoxide also enhanced SOD level in different tissues of animals.^{2,6,8,13,14,16,19,20,22,24,37,38,43,46,51,59,63}

Among the included studies, 21 assessed the effect of vanadium on CAT levels. In all of the studies, the CAT level significantly increased, except for two studies in which vanadium administration was not efficient^{8,46} and in 4 studies in which CAT level declined.^{16,42,45,52} In 6 studies,^{21,42,48,50,52,60} vanadyl sulfate administration significantly restored the altered enzymatic activity level of CAT to normal level. Sodium orthovanadate, vanadium-3-hydroxy flavone complex, macrocyclic binuclear oxovanadium (IV) complex, bis(maltolato) oxovanadium (IV), vanadium citrate, oxovanadium (IV) chelate were also effective in restoring CAT enzymatic activity level to near normal level in different tissues.^{6,14,16,19,21,22,24,37,45,46,51,59,63} However, in one study sodium orthovanadate administration was not effective in altering CAT level.8

Inflammatory biomarkers were assessed in 7 studies and significant decline was observed in all studies. Vanadyl sulfate supplementation significantly reduced TNF- α , IL-6, and hs-CRP.^{40,41,44,58,63} Vanadyl (IV)-ascorbate (VOAsc) complex and dioxidovanadium also reduced these inflammatory biomarkers.^{43,61} Caspase 3 level significantly decreased after oxovanadium (IV) complex and VOSO4 treatment.⁶⁶

Discussion

In the present systematic review, most of the studies claimed beneficial features for vanadium concerning inflammatory and oxidative stress biomarkers in their overall conclusion, despite treatment with different compounds of vanadium, doses, species, methods of administration, and length of intervention. In the included studies, impaired enzymes levels expressed as either decrease or increase, were accepted as oxidative stress. For example, in few studies in which antioxidant enzymes were enhanced in the

Saken et al, 1993 1	Sekar et al, 1990	+	;	+	•	+	+	•	?	+	?
Thompson MANNALL Image: 1996 Image: 1996 <thimage: 1996<="" th=""></thimage:>	H.Oster et al, 1993	•	;	•	+	+	+	•	;	•	+
1993 0		+	+	:	•	+	•	+	:	+	:
Gend et al., 2002 1 </td <td>1993</td> <td>+</td> <td>+</td> <td>•</td> <td>+</td> <td>•</td> <td>+</td> <td>•</td> <td>:</td> <td>•</td> <td>\$</td>	1993	+	+	•	+	•	+	•	:	•	\$
Cupta et al., 2004 P	Gupta and Baquer, 1998	+	+	:	•	+	•	+	+	-	:
Ranchandram et al, 2005 Image: Construction of the second sec				•		•	+		•		+
2004 T		-	-	-	-	-	-	+	-	-	+
Koyuhr, et al, 2005 Image: transmitted and trans	2004	+	+	•		+		•	-	_	+
Siddiqui et al, 2005 R. Wilksy et al, 2006 Imail and Yamardag, 2007 Imail and Yamardag, 2008 Imail and Yamardag, 20				•		+		•		-	+
R. Wilksy et al, 2007 Imail and Yunardag, 2006 Imail and Yunardag, 2007 Imail and Yunardag		+		-	+	-	+	-			+
Tunali and Yanardag. 1	en no anti-serve a la constante en	+			+			-			(
2006 (2) (2)	and the second second		-	-	-	-			-	-	+
2006 0		-	-	-	•	-	T	-	-	-	-
Shukla et al, 2007 + +	2006	-	-		-	+	-	+	-	-	+
Chareeb and Hussen, 2008				-	+		-		-		Ŧ
2008 			-	-				-	-	-	2
Yunardag et al, 2009 +				-	-	-	-	-	-		
Kurt et al, 2011 1	Yanardag et al, 2009					-					+
Kumar et al, 2012 + + 5 + + 5 <td< td=""><td>Kurt et al, 2011</td><td></td><td></td><td>-</td><td></td><td>-</td><td>-</td><td>-</td><td>?</td><td>_</td><td>+</td></td<>	Kurt et al, 2011			-		-	-	-	?	_	+
Sanchez-Gonzalez et al. 2013	Kumar et al, 2012			?		+	+	ě			+
All2 Imail and Yanardag.		-	-		+		-	+	?	?	+
2013 Image: Constraint of the second sec	Tunali and Yanardag,	-	-	-	-		-	-	?	+	+
Pillai et al, 2014 5 +		-	-	-	-			-		-	+
Sanchez-Gonzalez et al, 2014 Sun et al, 2014 Xie et al, 2014 Yilmaz-Ozden et al, 2014 Yilmaz-Ozden et al, 2014 Yilmaz-Ozden et al, 2014 Yilmaz-Ozden et al, 2014 Yilmaz-Ozden et al, 2014 Kumar et al, 2015 Liu et al, 2015 El-Karib et al, 2016 Ahmadi-Eslamloo et al, 2017 Bin-Jaliah et al, 2018 Sushko et al, 2018 Sushko et al, 2019 Sibiya et al, 2019 Bin-Jaliah et al, 2020 Liu et al, 2020 Tunai et al, 2020 Sinchez-González et al, 2021 Sinchez-González et al, 2017 Sinchez-González et al, 2018 Sushko et al, 2019 Sinchez-González et al, 2017 Sinchez-González et al, 2017 Sinchez-González et al, 2017 Sinchez-González et al, 2017 Sinchez-González et al, 2018 Sinchez-González et al, 2017 Sinchez-González et al, 2017 Sinchez-González et al, 2017 Sinchez-González et al, 2017 Sinchez-González et al, 2018	Pillai et al, 2014	•	-	-	-	-	-	-	-	-	+
2014 T	Sanchez-Gonzalez et al,	-	-	-	-	-	•	-		-	-
Xie et al, 2014 1	2014	_	_	-	-	-	-	-	-	-	+
Yilmaz-Ozden et al, 1		-	-	-	+	-	-	-	-	-	+
2014 1		-	-	•	-	-	+	•	-	-	+
2014 T	2014	-	-	-	-	+	•	•		-	+
Liu et al, 2015 •	2014	-	+	•	+	•	•	-	•	-	+
El-Karib et al, 2016 *		<u>;</u>	+	•	+	•	+	+	•	+	+
Ahmadi-Eslamloo et al, 2017 Imadi-Eslamloo et al, 2018 Imadi-Eslamloo et al, 2018 <t< td=""><td></td><td>+</td><td>+</td><td>•</td><td>+</td><td>+</td><td>+</td><td>•</td><td>•</td><td>?</td><td>+</td></t<>		+	+	•	+	+	+	•	•	?	+
2017 T		2	+	?	•	+	•	+	•	+	+
Espinosa-Zurutuza et al, 2018 • <t< td=""><td>2017</td><td>+</td><td>+</td><td>•</td><td>+</td><td>°)</td><td>•</td><td>+</td><td>•</td><td>+</td><td>•</td></t<>	2017	+	+	•	+	°)	•	+	•	+	•
2018 1	Bin-Jaliah et al, 2018	•	2	+	+	•	?	•	+	?	+
Sushko et al, 2018 +		+	+	•	+	•	+	•	:	+	+
Morsy et al, 2019 1	Samira et al, 2018	+	+	•	+	•	•	+	3	+	+
Sibiya et al, 2019 1	Sushko et al, 2018	+	+	•	+	•	+	•	2	:	+
Vijay et al, 2019 + + + + + - </td <td>Morsy et al, 2019</td> <td>+</td> <td>+</td> <td>•</td> <td>+</td> <td>•</td> <td>?</td> <td>+</td> <td>•</td> <td>?</td> <td>+</td>	Morsy et al, 2019	+	+	•	+	•	?	+	•	?	+
Bin-Jaliah et al, 2020 + + + + + + + * <td>Sibiya et al, 2019</td> <td>•</td> <td>?</td> <td>+</td> <td>?</td> <td>•</td> <td>?</td> <td>•</td> <td>2</td> <td>?</td> <td>+</td>	Sibiya et al, 2019	•	?	+	?	•	?	•	2	?	+
El-Shafey and El-Shafey and El-Sherbiny, 2020 Mbatha et al, 2020 Tunali et al, 2020 Sánchez-González et al, 2021	Vijay et al, 2019	+	+	+	•	•	?	•	2	+	•
Elsherbiny, 2020 Mbatha et al, 2020 Tunali et al, 2020 Sánchez-González et al, 2021 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	Bin-Jaliah et al, 2020	+	+	•	+	•	•	+	2	?	?
Mbatha et al, 2020 •	El-Shafey and	+	+	•	•	+	?	•	?	+	+
Sánchez-González et al, 2021 $ \begin{array}{c ccccccccccccccccccccccccccccccccccc$?	+	?	•	+	+	2	+	•
Sánchez-González et al,	Tunali et al, 2020	+	2	+	?	+			2	-	+
2021		_	-	-	-	-		+	-	-	+
sequence generation (Selection blas) Baseline characteristics (Selection blas) Allocation concealment (Selection blas) Aandom housing (Performance blas) Minding (Performance blas) Andom outcome assessment (Detection blas) tandom outcome assessment (Detection blas) candom outcome assessment (Detection blas) detective outcome cata (Attrition blas) effective outcome reporting (Reporting blas)	2021	-	•	-	•		-	•			
Sequence Baseline Alhoeatd Andom Bilnding Alhor or elective		e generation (Selection blas)	characteristics (Selection bias)	on concealment (Selection bias)	housing (Performance blas)	(Performance bias)	outcome assessment (Detection blas)	(Detection bias)	ete outcome data (Attrition blas)	outcome reporting (Reporting bias)	urces of blas (Other)
		duenc	seline	locati	nobn	nding	nobn	Inding	lqmoc	lectiv	her s(
· · · · · · · · · · · · · · · · · · ·		Seq	Bas	Alle	Raı	Blin	Ra	BIII	По	Sele	Oth

Figure 2. Risk of bias indicating studies' quality assessment at an individual level. (+) Low risk of bias. (-) High risk of bias. (?) Unclear risk of bias

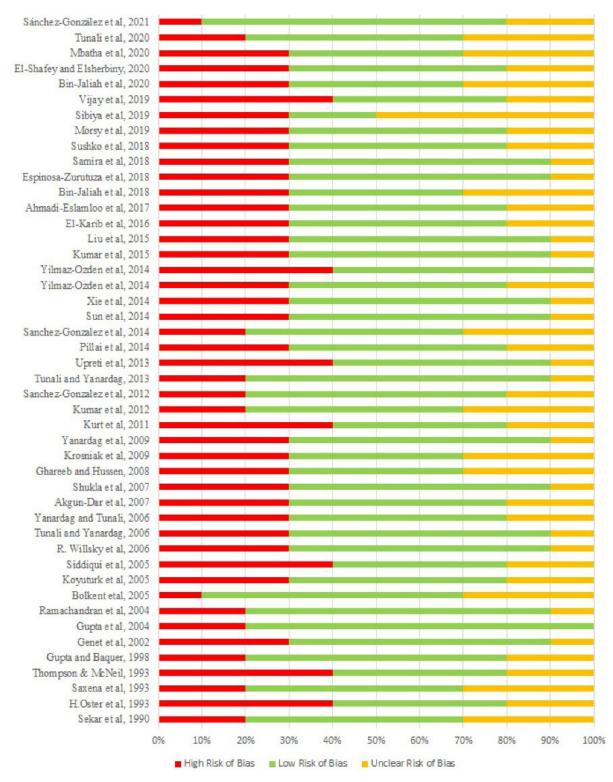


Figure 3. Risk of bias of included studies as percentages

cells for compensating an antioxidant defense during diabetes, vanadium administration was able to restore the number of enzymes impaired.^{42,53,54} Despite the observed beneficial impact of vanadium compounds in controlling inflammatory and oxidative stress biomarkers, there were few studies that indicated no significant effects.

In Saxena and colleagues' study, vanadate supplementation was useful in optimizing antioxidant enzymes of diabetes-induced animal models. This was mainly because vanadium in the form of vanadate was able to create different forms of free radicals, or distract the antioxidant system.⁴⁶ In Gupta and colleagues' study, no significant changes in the activity of CAT was observed after vanadium treatment. In the study of Krośniak et al, vanadium (V) peroxocomplexes treatment was not efficient regarding SOD level. This may be due to various oxidative positions and organic/inorganic ligands of vanadium (V) peroxocomplexes.¹³ The increased NAD(P) H: quinone-oxidoreductase-1 activity in diabetic rats may have decreased the formation of ROS, which would explain why no changes in SOD level were observed.⁴⁵ A recent study claimed that the administration of 1 mg/ day BMOV in diabetes-induced animal models was not efficient in declining inflammatory biomarkers.⁵⁶

As far as we know, the current systematic review was the first to assess the effect of vanadium on inflammatory and oxidative stress biomarkers in diabetes-induced animals. The last publications regarding vanadium and diabetes were mainly regarding glycemic factors.³⁰ Hence, several advantages could be mentioned for the present study. First, it included a high number of animal studies. Second, it assessed various outcomes. Third, it evaluated the effect of various organic and inorganic vanadium forms. Forth, it used the SYRCLE's risk of bias tool for evaluating the methodological quality of studies. However, few limitations can be mentioned for this systematic review, as below: (1) non-English studies were excluded; (2) gray literatures were not extracted during further searches (3) studies that supplemented vanadium along with insulin and/ or other compounds were also included.

Conclusion

The present systematic review reaffirmed that vanadium compounds in different doses and methods of administration were efficient in normalizing inflammatory and oxidative stress biomarkers in T2DM. Furthermore, addressing high-quality clinical trials for assessing the effectiveness of vanadium is encouraged.

Authors' contributions

FG and AO were involved in the concept of the manuscript; FG was responsible for writing the draft of the manuscript; MSA reviewed and edited the manuscript; and the final manuscript was read and approved by all of the authors.

Funding

The research protocol was funded by Student Research Committee, Tabriz University of Medical Sciences (grant number: 67836), Tabriz, Iran.

Ethical approval

Not applicable.

Competing interests

No conflict of interest was reported.

Disclaimer

The authors claim that no part of this paper is copied from other sources.

Supplementary Files

Supplementary file 1 contains Table S1.

References

- Burgos-Morón E, Abad-Jiménez Z, Marañón AM, Iannantuoni F, Escribano-López I, López-Domènech S, et al. Relationship between oxidative stress, ER stress, and inflammation in type 2 diabetes: the battle continues. J Clin Med. 2019;8(9):1385. doi: 10.3390/jcm8091385.
- 2. El-Shafey ES, Elsherbiny ES. The role of apoptosis and

autophagy in the insulin-enhancing activity of oxovanadium (IV) bipyridine complex in streptozotocin-induced diabetic mice. Biometals. 2020;33(2-3):123-35. doi: 10.1007/s10534-020-00237-1.

- 3. Abeeleh MA, Ismail ZB, Alzaben KR, Abu-Halaweh SA, Al-Essa MK, Abuabeeleh J, et al. Induction of diabetes mellitus in rats using intraperitoneal streptozotocin: a comparison between 2 strains of rats. Eur J Sci Res. 2009;32(3):398-402.
- Chistiakov DA, Bobryshev YV, Orekhov AN. Macrophagemediated cholesterol handling in atherosclerosis. J Cell Mol Med. 2016;20(1):17-28. doi: 10.1111/jcmm.12689.
- Yuan T, Yang T, Chen H, Fu D, Hu Y, Wang J, et al. New insights into oxidative stress and inflammation during diabetes mellitusaccelerated atherosclerosis. Redox Biol. 2019;20:247-60. doi: 10.1016/j.redox.2018.09.025.
- 6. Kumar P, Taha A, Kale RK, McLean P, Baquer NZ. Beneficial effects of *Trigonella foenum graecum* and sodium orthovanadate on metabolic parameters in experimental diabetes. Cell Biochem Funct. 2012;30(6):464-73. doi: 10.1002/cbf.2819.
- Treviño S, Diaz A. Vanadium and insulin: partners in metabolic regulation. J Inorg Biochem. 2020;208:111094. doi: 10.1016/j.jinorgbio.2020.111094.
- Gupta BL, Baquer NZ. Hexokinase, glucose-6-phosphate dehydrogenase and antioxidant enzymes in diabetic reticulocytes: effects of insulin and vanadate. Biochem Mol Biol Int. 1998;46(6):1145-52. doi: 10.1080/15216549800204702.
- Ścibior A, Zaporowska H, Wolińska A, Ostrowski J. Antioxidant enzyme activity and lipid peroxidation in the blood of rats cotreated with vanadium (V+5) and chromium (Cr+3). Cell Biol Toxicol. 2010;26(6):509-26. doi: 10.1007/s10565-010-9160-8.
- Soussi A, Croute F, Soleilhavoup JP, Kammoun A, El-Feki A. [Impact of green tea on oxidative stress induced by ammonium metavanadate exposure in male rats]. C R Biol. 2006;329(10):775-84. doi: 10.1016/j.crvi.2006.07.004.
- 11. Gupta BL, Preet A, Baquer NZ. Protective effects of sodium orthovanadate in diabetic reticulocytes and ageing red blood cells of Wistar rats. J Biosci. 2004;29(1):73-9. doi: 10.1007/bf02702564.
- Koyuturk M, Tunali S, Bolkent S, Yanardag R. Effects of vanadyl sulfate on liver of streptozotocin-induced diabetic rats. Biol Trace Elem Res. 2005;104(3):233-47. doi: 10.1385/ bter:104:3:233.
- Krośniak M, Gawlik M, Gryboś R. Effect of vanadium complexes and insulin administered simultaneously for oxidative stress in STZ diabetic rats. Bull Vet Inst Pulawy. 2009;53(3):535-40. doi: 10.1007/s12011-013-9688-6.
- Ramachandran B, Ravi K, Narayanan V, Kandaswamy M, Subramanian S. Protective effect of macrocyclic binuclear oxovanadium complex on oxidative stress in pancreas of streptozotocin induced diabetic rats. Chem Biol Interact. 2004;149(1):9-21. doi: 10.1016/j.cbi.2004.06.007.
- Yadav UC, Moorthy K, Baquer NZ. Effects of sodiumorthovanadate and *Trigonella foenum-graecum* seeds on hepatic and renal lipogenic enzymes and lipid profile during alloxan diabetes. J Biosci. 2004;29(1):81-91. doi: 10.1007/ bf02702565.
- Shukla R, Padhye S, Modak M, Ghaskadbi SS, Bhonde RR. Bis (quercetinato) oxovanadium IV reverses metabolic changes in streptozotocin-induced diabetic mice. Rev Diabet Stud. 2007;4(1):33-43. doi: 10.1900/rds.2007.4.33.
- Thompson KH, McNeill JH. Effect of vanadyl sulfate feeding on susceptibility to peroxidative change in diabetic rats. Res Commun Chem Pathol Pharmacol. 1993;80(2):187-200.
- Bolkent S, Bolkent S, Yanardag R, Tunali S. Protective effect of vanadyl sulfate on the pancreas of streptozotocin-induced diabetic rats. Diabetes Res Clin Pract. 2005;70(2):103-9. doi:

10.1016/j.diabres.2005.02.003.

- 19. Upreti J, Ali S, Basir SF. Effect of lower doses of vanadate in combination with *Azadirachta indica* leaf extract on hepatic and renal antioxidant enzymes in streptozotocin-induced diabetic rats. Biol Trace Elem Res. 2013;156(1-3):202-9. doi: 10.1007/s12011-013-9827-0.
- Vijay K, Suresh R, Loganathasamy K, Narayanan V, Pratheepa K, Venkataraman K, et al. Antioxidant status in STZ-induced diabetic rats treated with vanadium pentoxide nanoparticles. Indian J Anim Res. 2019;53(12):1594-8. doi: 10.18805/ijar.B-3709.
- Xie M, Chen D, Zhang F, Willsky GR, Crans DC, Ding W. Effects of vanadium (III, IV, V)-chlorodipicolinate on glycolysis and antioxidant status in the liver of STZ-induced diabetic rats. J Inorg Biochem. 2014;136:47-56. doi: 10.1016/j. jinorgbio.2014.03.011.
- Genet S, Kale RK, Baquer NZ. Alterations in antioxidant enzymes and oxidative damage in experimental diabetic rat tissues: effect of vanadate and fenugreek (*Trigonellafoenum* graecum). Mol Cell Biochem. 2002;236(1-2):7-12. doi: 10.1023/a:1016103131408.
- Ahmadi-Eslamloo H, Dehghani GA, Moosavi SMS. Longterm treatment of diabetic rats with vanadyl sulfate or insulin attenuate acute focal cerebral ischemia/reperfusion injury via their antiglycemic effect. Metab Brain Dis. 2018;33(1):225-35. doi: 10.1007/s11011-017-0153-7.
- Sushko O, Ponkalo L. Glutathione status in rat's liver experimentally induced under influence chromium citrate. Ukr Biochem J. 2018;90(6):141. doi: 10.15407/ubj90.06.
- Chandra AK, Ghosh R, Chatterjee A, Sarkar M. Effects of vanadate on male rat reproductive tract histology, oxidative stress markers and androgenic enzyme activities. J Inorg Biochem. 2007;101(6):944-56. doi: 10.1016/j. jinorgbio.2007.03.003.
- Srivastava A.K, Mehdi M. Z. Insulino-mimetic and anti-diabetic effects of vanadium compounds. Diabet. Med. 2005;22(1):2-13. doi: 10.1111/j.1464-5491.2004.01381.x.
- Willsky GR, Chi LH, Liang Y, Gaile DP, Hu Z, Crans DC. Diabetes-altered gene expression in rat skeletal muscle corrected by oral administration of vanadyl sulfate. Physiol Genomics. 2006;26(3):192-201. doi: 10.1152/ physiolgenomics.00196.2005.
- Ramachandran B, Ravi K, Narayanan V, Kandaswamy M, Subramanian S. Effect of macrocyclic binuclear oxovanadium complex on tissue defense system in streptozotocin-induced diabetic rats. Clin Chim Acta. 2004;345(1-2):141-50. doi: 10.1016/j.cccn.2004.03.014.
- Smith DM, Pickering RM, Lewith GT. A systematic review of vanadium oral supplements for glycaemic control in type 2 diabetes mellitus. QJM. 2008;101(5):351-8. doi: 10.1093/ gjmed/hcn003.
- Berhan A, Habtewolde A. Effects of vanadium compounds on glycemic control in type 2 diabetes mellitus: a metaanalysis of comparative study on rats. Int J Pharm Sci Res. 2012;3(10):3717-24.
- Peters JL, Sutton AJ, Jones DR, Rushton L, Abrams KR. A systematic review of systematic reviews and metaanalyses of animal experiments with guidelines for reporting. J Environ Sci Health B. 2006;41(7):1245-58. doi: 10.1080/03601230600857130.
- 32. Moher D, Liberati A, Tetzlaff J, Altman DG, Altman D, Antes G, et al. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement (Chinese edition). Chin J Integr Med. 2009;7(9):889-96. doi: 10.3736/jcim20090918.
- Leenaars M, Hooijmans CR, van Veggel N, ter Riet G, Leeflang M, Hooft L, et al. A step-by-step guide to systematically identify all relevant animal studies. Lab Anim. 2012;46(1):24-31. doi: 10.1258/la.2011.011087.

- 34. Hooijmans CR, Tillema A, Leenaars M, Ritskes-Hoitinga M. Enhancing search efficiency by means of a search filter for finding all studies on animal experimentation in PubMed. Lab Anim. 2010;44(3):170-5. doi: 10.1258/la.2010.009117.
- Higgins JPT, Thomas J, Chandler J, Cumpston M, Li T, Page MJ, et al. Cochrane Handbook for Systematic Reviews of Interventions. Hoboken: Wiley; 2019. doi: 10.1002/9781119536604.
- Hooijmans CR, Rovers MM, de Vries RB, Leenaars M, Ritskes-Hoitinga M, Langendam MW. SYRCLE's risk of bias tool for animal studies. BMC Med Res Methodol. 2014;14:43. doi: 10.1186/1471-2288-14-43.
- Sekar N, Kanthasamy A, William S, Balasubramaniyan N, Govindasamy S. Antioxidant effect of vanadate on experimental diabetic rats. Acta Diabetol Lat. 1990;27(4):285-93. doi: 10.1007/bf02580932.
- Mbatha B, Khathi A, Sibiya N, Booysen I, Mangundu P, Ngubane P. Cardio-protective effects of a dioxidovanadium (V) complex in male Sprague-Dawley rats with streptozotocininduced diabetes. Biometals. 2021;34(1):161-73. doi: 10.1007/s10534-020-00270-0.
- 39. Akgün-Dar K, Bolkent S, Yanardag R, Tunali S. Vanadyl sulfate protects against streptozotocin-induced morphological and biochemical changes in rat aorta. Cell Biochem Funct. 2007;25(6):603-9. doi: 10.1002/cbf.1354.
- Bin-Jaliah I, Morsy MD, Al-Ani B, Eid RA, Haidara MA. [Vanadium inhibits type 2 diabetes mellitus-induced aortic ultrastructural alterations associated with the inhibition of dyslipidemia and biomarkers of inflammation in rats]. Int J Morphol. 2020;38(1):215-21. doi: 10.4067/s0717-95022020000100215.
- 41. El Karib AO, Al-Ani B, Al-Hashem F, Dallak M, Bin-Jaliah I, El-Gamal B, et al. Insulin and vanadium protect against osteoarthritis development secondary to diabetes mellitus in rats. Arch Physiol Biochem. 2016;122(3):148-54. doi: 10.3109/13813455.2016.1159698.
- 42. Kurt O, Ozden TY, Ozsoy N, Tunali S, Can A, Akev N, et al. Influence of vanadium supplementation on oxidative stress factors in the muscle of STZ-diabetic rats. Biometals. 2011;24(5):943-9. doi: 10.1007/s10534-011-9452-3.
- Liu Y, Xu J, Guo Y, Xue Y, Wang J, Xue C. Ameliorative effect of vanadyl (IV)-ascorbate complex on high-fat high-sucrose diet-induced hyperglycemia, insulin resistance, and oxidative stress in mice. J Trace Elem Med Biol. 2015;32:155-61. doi: 10.1016/j.jtemb.2015.07.007.
- 44. Morsy MD, Bin-Jaliah I, Bashir SO, Shatoor A, Haidara MA. The impact of concomitant administration of vanadium and insulin on endothelial dysfunction markers (PAI-1 and ET-1) in type 1 diabetic rats. Arch Physiol Biochem. 2021;127(1):20-7. doi: 10.1080/13813455.2019.1573840.
- Sánchez-González C, López-Chaves C, Trenzado CE, Aranda P, López-Jurado M, Gómez-Aracena J, et al. Changes in iron metabolism and oxidative status in STZ-induced diabetic rats treated with bis (maltolato) oxovanadium (IV) as an antidiabetic agent. ScientificWorldJournal. 2014;2014:706074. doi: 10.1155/2014/706074.
- Saxena AK, Srivastava P, Kale RK, Baquer NZ. Impaired antioxidant status in diabetic rat liver. Effect of vanadate. Biochem Pharmacol. 1993;45(3):539-42. doi: 10.1016/0006-2952(93)90124-f.
- 47. Sun L, Shi DJ, Gao XC, Mi SY, Yu Y, Han Q. The protective effect of vanadium against diabetic cataracts in diabetic rat model. Biol Trace Elem Res. 2014;158(2):219-23. doi: 10.1007/s12011-014-9925-7.
- Tunali S, Peksel A, Arisan I, Yanardağ R. Study of the beneficial effect of vanadium sulfate on the liver of experimental diabetic rats. Istanbul J Pharm. 2020;50(3):211-5. doi: 10.26650/ IstanbulJPharm.2020.0065.

- 49. Tunali S, Yanardag R. Effect of vanadyl sulfate on the status of lipid parameters and on stomach and spleen tissues of streptozotocin-induced diabetic rats. Pharmacol Res. 2006;53(3):271-7. doi: 10.1016/j.phrs.2005.12.004.
- Tunali S, Yanardag R. Protective effect of vanadyl sulfate on skin injury in streptozotocin-induced diabetic rats. Hum ExpToxicol. 2013;32(11):1206-12. doi: 10.1177/0960327113478445.
- Yanardag R, Demirci TB, Ulküseven B, Bolkent S, Tunali S, Bolkent S. Synthesis, characterization and antidiabetic properties of N1-2,4-dihydroxybenzylidene-N4-2-hydroxybenzylidene-S-methyl-thiosemicarbazidato-oxovanadium (IV). Eur J Med Chem. 2009;44(2):818-26. doi: 10.1016/j.ejmech.2008.04.023.
- 52. Yanardag R, Tunali S. Vanadyl sulfate administration protects the streptozotocin-induced oxidative damage to brain tissue in rats. Mol Cell Biochem. 2006;286(1-2):153-9. doi: 10.1007/ s11010-005-9107-1.
- 53. Yilmaz-Ozden T, Kurt-Sirin O, Tunali S, Akev N, Can A, Yanardag R. Ameliorative effect of vanadium on oxidative stress in stomach tissue of diabetic rats. Bosn J Basic Med Sci. 2014;14(2):105-9. doi: 10.17305/bjbms.2014.2273.
- 54. Yilmaz-Ozden T, Kurt-Sirin O, Tunali S, Akev N, Can A, Yanardag R. Effect of oral vanadium supplementation on oxidative stress factors in the lung tissue of diabetic rats. Trace Elements Electrolytes. 2014;31(2):48-52. doi: 10.5414/ tex01317.
- Sanchez-Gonzalez C, Bermudez-Peña C, Trenzado CE, Goenaga-Infante H, Montes-Bayon M, Sanz-Medel A, et al. Changes in the antioxidant defence and in selenium concentration in tissues of vanadium exposed rats. Metallomics. 2012;4(8):814-9. doi: 10.1039/c2mt20066j.
- Sánchez-González C, Rivas-García L, Rodríguez-Nogales A, Algieri F, Gálvez J, Aranda P, et al. Vanadium decreases hepcidin mRNA gene expression in STZ-induced diabetic rats, improving the anemic state. Nutrients. 2021;13(4):1256. doi: 10.3390/nu13041256.
- Willsky GR, Chi LH, Liang Y, Gaile DP, Hu Z, Crans DC. Diabetes-altered gene expression in rat skeletal muscle corrected by oral administration of vanadyl sulfate. Physiol Genomics. 2006;26(3):192-201. doi: 10.1152/ physiolgenomics.00196.2005.
- 58. Bin-Jaliah I, Sakr HF, Morsy MD, Dallak M, Haidara MA. Modulatory effect of concomitant administration of insulin

and vanadium on inflammatory biomarkers in type 2 diabetic rats: role of adiponectin. Chin J Physiol. 2018;61(1):42-9. doi: 10.4077/cjp.2018.bag523.

- 59. Pillai SI, Subramanian SP, Kandaswamy M. Antidyslipidemic effect of a novel vanadium-3-hydroxy flavone complex in streptozotocin-induced experimental diabetes in rats. Biomed Prev Nutr 2014;4(2):189-93. doi: 10.1016/j. bionut.2013.04.004.
- 60. Samira M, Mounira T, Kamel K, Yacoubi MT, Ben Rhouma K, Sakly M, et al. Hepatotoxicity of vanadyl sulfate in nondiabetic and streptozotocin-induced diabetic rats. Can J Physiol Pharmacol. 2018;96(11):1076-83. doi: 10.1139/cjpp-2018-0255.
- 61. Sibiya S, Msibi B, Khathi A, Sibiya N, Booysen I, Ngubane P. The effect of dioxidovanadium complex (V) on hepatic function in streptozotocin-induced diabetic rats. Can J Physiol Pharmacol. 2019;97(12):1169-75. doi: 10.1139/cjpp-2019-0369.
- 62. Ghareeb DA, Hussen HM. Vanadium improves brain acetylcholinesterase activity on early stage alloxan-diabetic rats. Neurosci Lett. 2008;436(1):44-7. doi: 10.1016/j. neulet.2008.02.073.
- 63. Siddiqui MR, Taha A, Moorthy K, Hussain ME, Basir SF, Baquer NZ. Amelioration of altered antioxidant status and membrane linked functions by vanadium and *Trigonella* in alloxan diabetic rat brains. J Biosci. 2005;30(4):483-90. doi: 10.1007/bf02703722.
- 64. Kumar P, Taha A, Kumar N, Kumar V, Baquer NZ. Sodium orthovanadate and *Trigonella foenum graecum* prevents neuronal parameters decline and impaired glucose homeostasis in alloxan diabetic rats. Prague Med Rep. 2015;116(2):122-38. doi: 10.14712/23362936.2015.51.
- 65. Oster MH, Llobet JM, Domingo JL, German JB, Keen CL. Vanadium treatment of diabetic Sprague-Dawley rats results in tissue vanadium accumulation and prooxidant effects. Toxicology. 1993;83(1-3):115-30. doi: 10.1016/0300-483x(93)90096-b.
- El-Shafey ES, Elsherbiny ES. The role of apoptosis and autophagy in the insulin-enhancing activity of oxovanadium (IV) bipyridine complex in streptozotocin-induced diabetic mice. Biometals. 2020;33(2-3):123-35. doi: 10.1007/s10534-020-00237-1.